

GENOTYPING OF GIARDIA LAMBLIA AND CRYPTOSPORIDIUM SPP

FROM PANAMANIAN CHILDREN UNDER FIVE YEARS OLD.

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INTRODUCTION

Giardia lamblia and Cryptosporidium spp are enteropathogens parasites that can cause gastrointestinal disease and nutritional deficiency in children. These two pathogenic protozoa are present with significant frequencies in rural and suburban communities in Panama, Central America. In some regions of the country it is possible to find giardiasis prevalence between 10-50%. The situation for cryptosporidiosis is less clear. In previous studies we have detected a prevalence between 2-15%. However, information on their genetic characteristics, distribution, and role in human disease in Panama is limited. The molecular characterization of species and genotypes of Cryptosporidium and Giardia is also necessary for identifying and assessing zoonotic transmission. The present study represents the first contribution in Panama concerning G. lamblia and Cryptosporidium spp. genotypes circulating in children form different areas of the country.



Figure 2. RSA I digestion of the PCR-multiplex products (Figure 1). Lane 1, 100-bp DNA ladder; lanes 4 - 10 correspond to subgenotype A1 (437 bp); lanes 5, 8 and 9 to subgenotype A2. The product of 140 bp, corresponding to Assemblage B was not digested.

OBJECTIVE

To genotype *Giardia* and *Cryptosporidium* isolates from infected Panamanian children under five year of age.

METHODOLOGY

We analyzed the genetic diversity and geographic distribution of both protozoa from infected children younger than five years. Stool samples were taken from 1,840 diarrheic and non-diarrheic children from eight different districts in Panama: Panama, La Chorrera, Cañazas, Santa Fe, Changuinola, Panama Este, Ipeti-Choco and San Felix (Figure 1).



Figure 1. PCR-Multiplex analysis using the tpi gene of *G. lamblia.* Lane 1,100-bp DNA ladder; lane 2, 4, 5 and 10 Assemblage A (476 bp); lanes 3, 6 and 7 Assemblage B (140 bp); lanes 8 and 9 are mixed infections.



Figure 3. Identification of *Giardia* Assemblages (A and B) and subgenotypes (A1 and A2) in infected Panamanian children less than five years of age from different regions of the country.

Table I. *Cryptosporidium* genotypes and subtypes from different regions of Panama

SPECIES	REGION	PANAMA	CHORRERA	CHANGUINOLA	SANTA FE	CAÑAZAS	SAN FELIX	CHEPO
Total	N=24	N=3	N=6	N=5	N=2	N=2	N=5	N=1
C. hominis	n=16	1	5*	5	1*	1	3*	
subtype family	Ie	1	3	4			1	
(12/16)	Id			1		1		
	Ib		1					
C. parvum	n=4							1*
subtype family	IIa		1					
(3/4)	IIc	2						

Fig 1: Different Regions of Panama G. studied *lamblia* and for *Cryptosporidium* spp. in children under five years old.

C. meleagridis	n=3		1		2	
C.canis	n=1			1		

ng 108 Changuinola B051 Chorrera (379 San Félix J493091 Brazil J493534 2 Brazi F108865 Australi Cryptosporidium hominis 570922 China J493070 Kenva J493536 Brazil -044 Santa Fe 159 Changuinola Cryptosporidium parvum Cryptosporidium meleagridis 019 San Félix F003 Santa Fé ABORATOR AF329185 Perú FX183 San Félix

Figure 5. Phylogenetic relationships of the examined isolates with other Cryptosporidium genotypes as inferred by Neighbor-joining analysis, based on the nucleotide sequence of SSU rRNA. The name of the isolates accession numbers in GenBAnk are shown in red.

Cysts of G. lamblia were microscopically detected using the formalin-acetate concentration procedure. For oocysts detection a Kinyoun staining procedure was used. DNA was extracted only from positive samples. (Fig.2) G. lamblia genotyping was performed using a PCR-RFLP analysis based on the polymorphisms of the *tpi* gene. For Cryptosporidium spp, the SSU rRNA gene was used as molecular marker. Cryptosporidium identification was confirmed by direct sequence analysis. Further genetic diversity within *C. parvum* and *C. hominis* samples were assessed by sequence analysis of the GP60 gene.



Fig.2: a. Cysts of *G. lamblia* observed in stool samples. b. Oocysts of Cryptosporidium sp. c .Purification of *Cryptosporidium* samples for genotype analysis.

RESULTS

Of the evaluated samples, 341 (18.5%) presented G. lamblia cysts and 93 (5.1%) presented Cryptosporidium spp oocysts. DNA was extracted from all of these samples. Molecular diagnosis and characterization was only possible in 132 Giardia and 24 Cryptosporidium positive samples. Genetic analysis of the tpi gene revealed that of the 132 genotyped Giardia samples 25 (19%) belonged to assemblage A, 99 (75.0%) belonged to assemblage B and the remaining 8 (6.1%) were mixed infections (Figures 2 and 3). A preliminary subtyping analysis of assemblage A samples demonstrated that the AII type was nearly nine times more frequent than AI type (Figure 4). Genotyping of 24 Cryptosporidium isolates from Panamanian children identified four Cryptosporidium species: 62.5% (15/24) were C. hominis, 21.0% (5/24) *C. parvum*, 12.5% (3/24) *C. meleagridis* and 4.2% (1/24) *C. canis*. (Table 1, Figure 5). The GP60 gen was sequenced from 15 samples. For C. hominis (n =12), 9 belong to the Ie allele, 2 for Id, and 1 for Ib. For *C. parvum* (n =3), two samples were IIc and one IIa.

CONCLUSIONS

G. lamblia and Cryptosporidium spp. are important protozoan infections in children from different regions of Panama. G. lamblia assemblage B (75%) is the main genotype and Cryptosporidium hominis (62.5%) the predominant species circulating in different regions of Panama. In addition, other potential zoonotic Cryptosporidium as C. meleagridis (12.5%) and C. canis (4.2%) were detected. C. hominis Ie and C. parvum IIc were the predominant subtypes circulating in Panama. □ Further studies of human and animal isolates are required to evaluate the molecular epidemiology of G. lamblia and Cryptosporidium spp. in Panama. This information is important for the development of systematic prevention and control measures for these enteroparasitic infections.